

**Gli elementi chiave da acquisire per
proteggere i pazienti fragili**

**Antibiotico resistenza ed infezioni
correlate all'assistenza**

26 NOVEMBRE - SALA MINERVA

PAOLA BERNASCHI



UOC Microbiologia e Diagnostica di Immunologia
Ospedale Pediatrico Bambino Gesù (IRCCS)
Roma

Chi sono le persone “fragili” e perché sono più a rischio se contraggono malattie infettive?

Tutti coloro che presentano condizioni morbose che espongono a un rischio maggiore di contrarre malattie infettive invasive e sviluppare in tal caso complicanze gravi



Severe newborn infections—including sepsis, meningitis, and **pneumonia**—are a major cause of morbidity and mortality among newborns in low- and middle-income countries.

Recent analyses estimate that globally, approximately 400,000 newborns die each year as a result of severe infections, over one-sixth of the total burden of newborn deaths. Most of these deaths could be averted through:

1. preventive measures, such as improving hygiene practices, and
2. timely and appropriate care for sick newborns.

17 percentage of neonatal deaths caused by severe bacterial infections

400K number of newborns who die each year as a result of severe infections

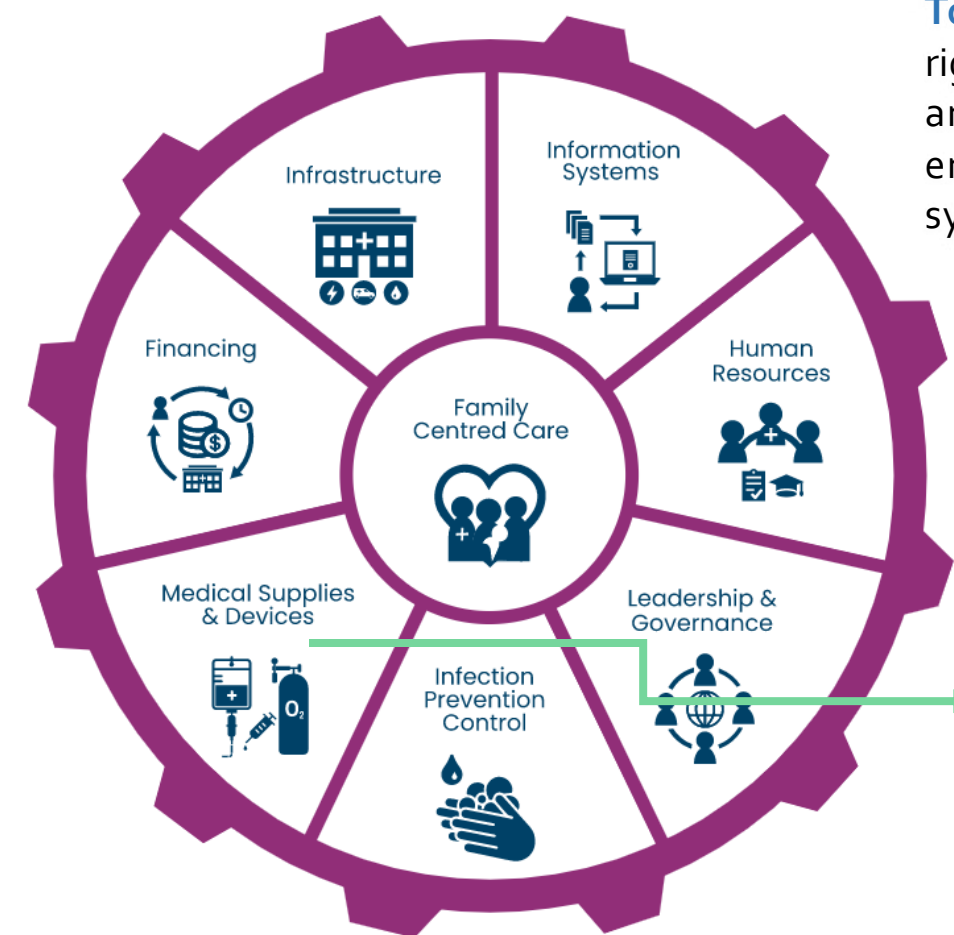
8 percentage of neonatal deaths caused by pneumonia

I bambini appena nati sono soggetti fragili perché devono ancora sviluppare il sistema immunitario, quindi più esposti per esempio al rischio di infezioni respiratorie e sistemiche

Date of update: 17 September 2022 

L'importanza della gestione clinica per la diagnostica microbiologica

To deliver high quality care for every small and sick newborn, we need to have the right space, the right people and the right supplies, backed by supportive leadership and data for action. This requires a package of care with families at the center ensuring zero separation between parents and their newborns. A whole health systems, country-led approach is required –which is what this toolkit aims to achieve.



Medical Supplies & Devices



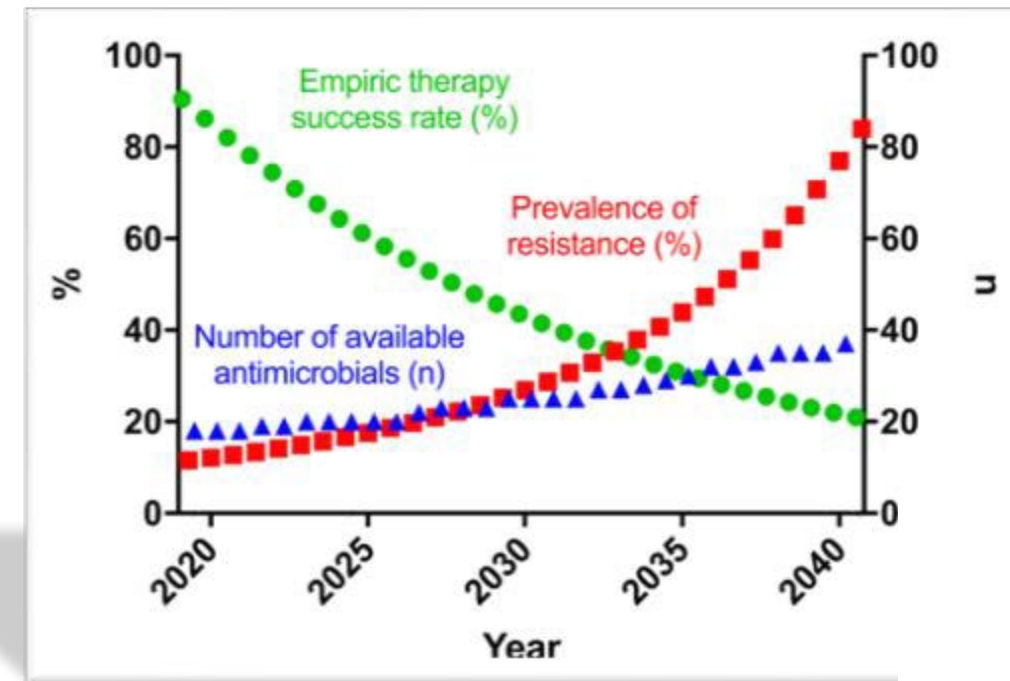
Laboratory Medicine

It is estimated that 70% of medical decisions (in high-resource settings) are based on clinical laboratory tests. As a result, diagnostic testing is critical to saving the lives of many newborns. Laboratory medicine provides meaningful, accurate results to diagnose conditions, follow-up and monitor treatments

Clinical laboratories are key to providing highly reliable diagnostic data to inform clinicians on the best medical care they should provide to the newborn. Management and improvement of quality in laboratories over time are essential to achieve this goal

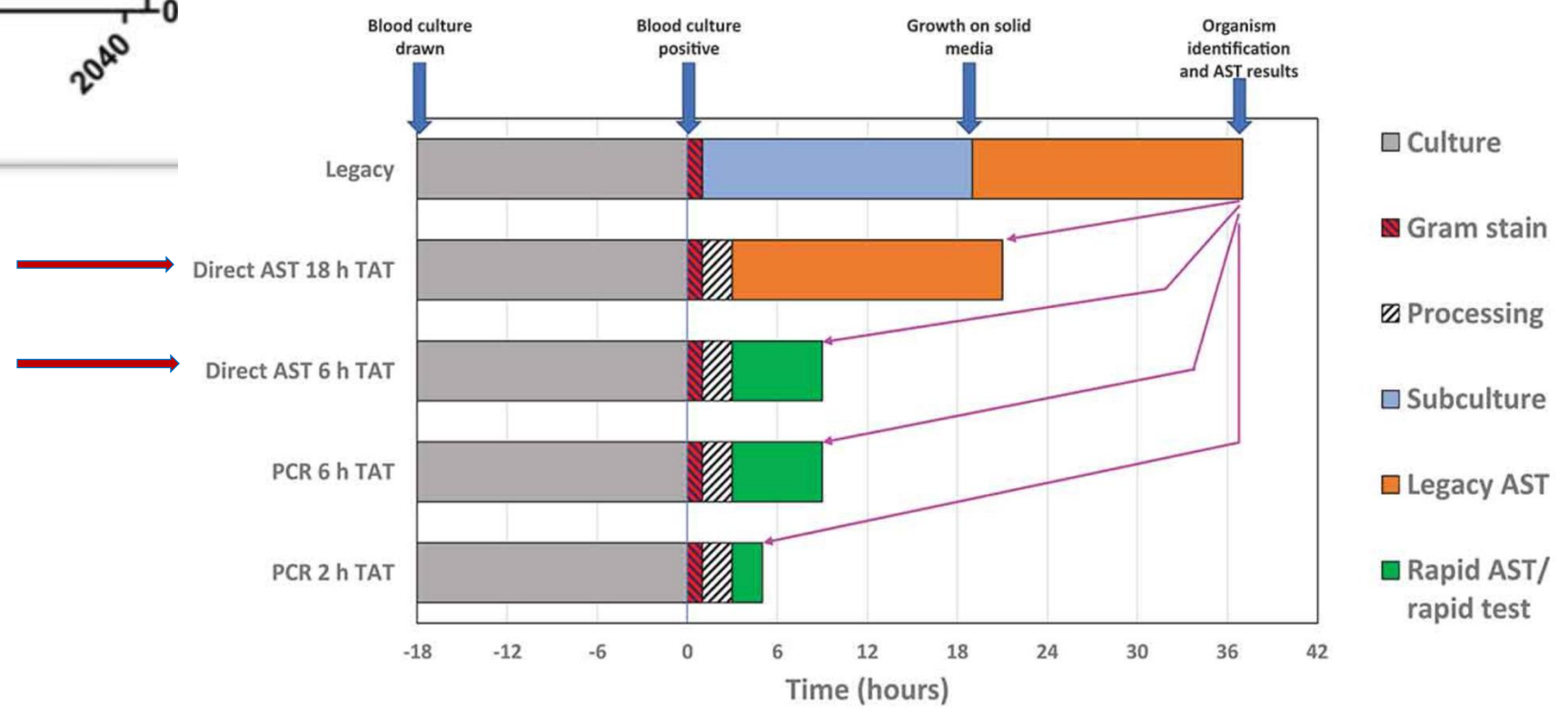
Date of update: 17 April 2022



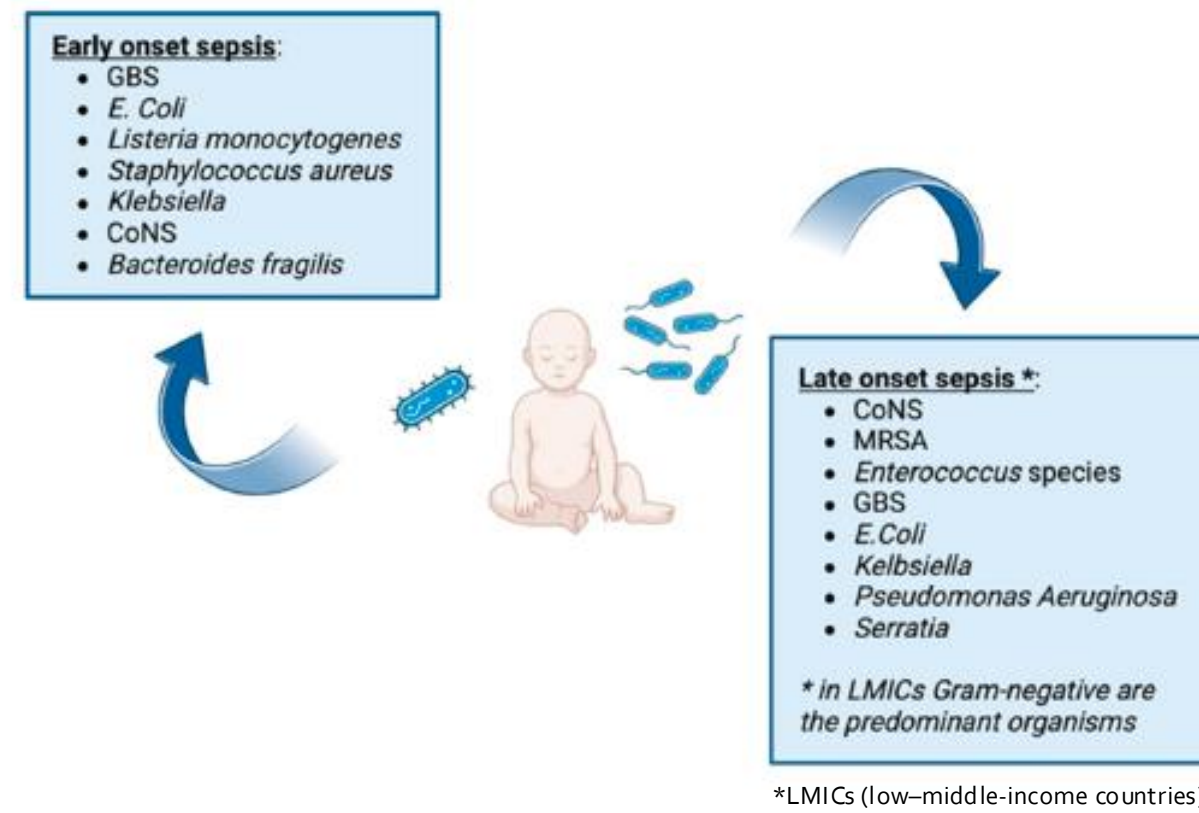


Loss of reliable empiric therapy

Timeline illustrating timing of blood culture results by legacy and rapid methods



Kenneth P. Smith et al. Clinics in Laboratory Medicine 2019
 Michael R. Jacobs et al. Expert Review of Molecular Diagnostics 2021



Major bacterial species associated with neonatal sepsis

Antibiotics 2024, 13, 250. <https://doi.org/10.3390/antibiotics13030250>



Review
An Overview of Antibiotic Therapy for Early- and Late-Onset Neonatal Sepsis: Current Strategies and Future Prospects

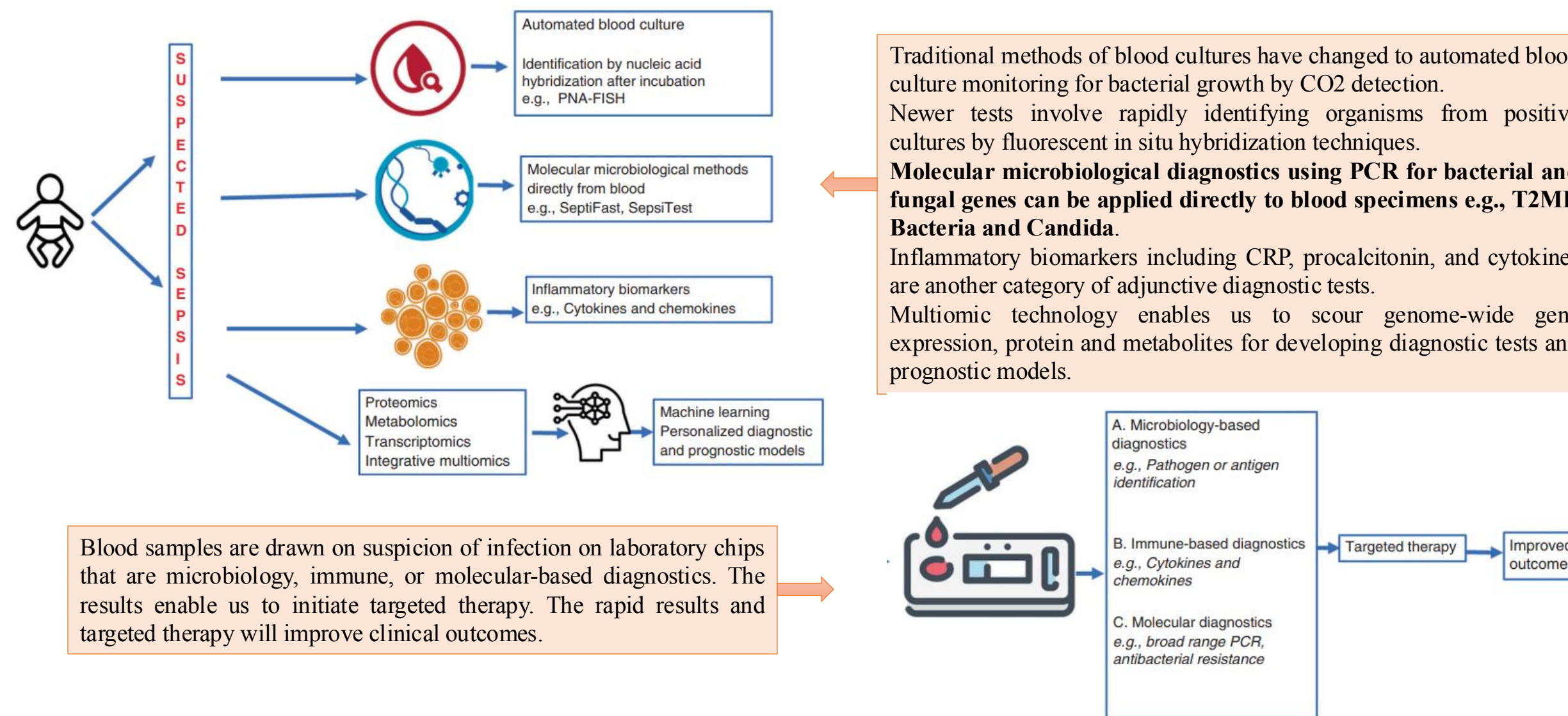
Giovanni Boscarino ¹, Rossana Romano ¹, Carlotta Iotti ¹, Francesca Tegoni ¹, Serafina Perrone ² and Susanna Esposito ^{1,*}

Empirical Antimicrobial Policies	Indication
β-lactam antibiotic + Aminoglycoside (Gentamicin)	Gram-positive and Gram-negative agents; this should be used in neonates non-colonized with MRSA to offer anti-staphylococcal coverage
β-lactam antibiotic + Aminoglycoside (Amikacin)	More resistant Gram-negative and some Gram-positive bacteria (i.e., <i>Staphylococcus aureus</i>); this could replace Gentamicin in selected cases (higher-risk preterm neonates or neonates with severe disease)
Glycopeptide + Aminoglycoside	Empiric Gram-positive and Gram-negative coverage; confirmed CoNS and MRSA
Piperacillin + Tazobactam or Ampicillin + Sulbactam	In combination or in alternative to aminoglycoside; Gram-positive and Gram-negative beta-lactamase-producing bacteria
Third- or fourth-generation Cephalosporin	In addition to empiric regimen; for severe penicillin-resistant Gram-negative sepsis or Gram-negative meningitis (no Ceftriaxone)
Carbapenems	ESBL and AmpC chromosomal beta-lactamase-producing Gram-negative; bacterial meningitis
Colistin	CRO

Common antibiotics for empirical therapy in suspected neonatal sepsis.

REVIEW ARTICLE

Diagnosis of neonatal sepsis: the past, present and future



Celik et al., *Pediatr Res.* 2022

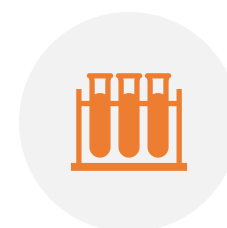
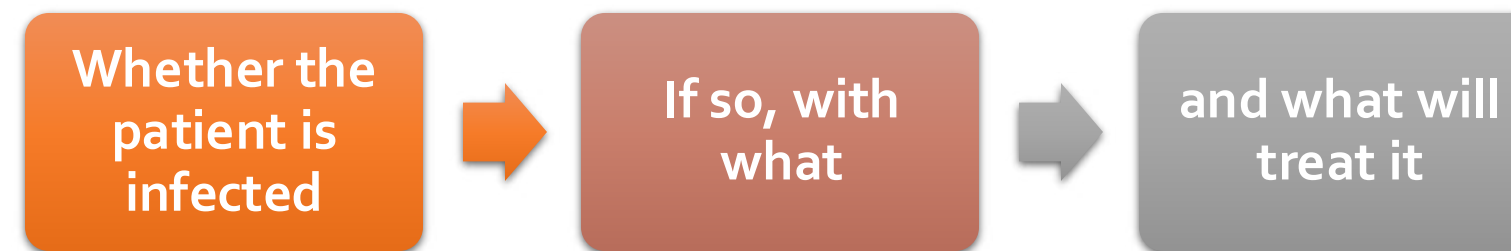
Bartlett RC. 1974 *Medical microbiology: quality cost and clinical relevance*. Wiley, New York, NY

“Just because you can does not necessarily mean you should”



“More practical, economical, clinically meaningful approach”

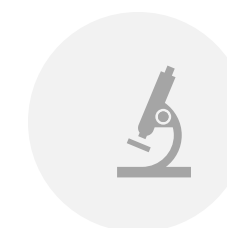
Clinicians seek three basic truths:



Guide the proper use of tests and the flow of results



Help optimize empiric antibiotic prescribing by creating a facility **cumulative antibiotic resistance report or antibiogram**

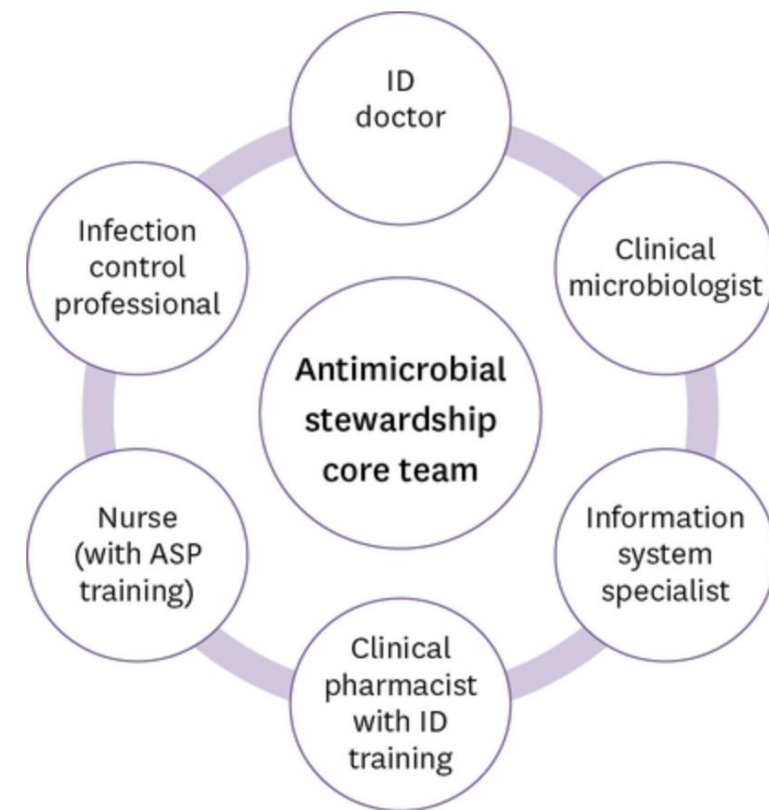


Microbiology labs and stewardship programs can work together to optimize the use of **rapid diagnostic tests** and the communication of results

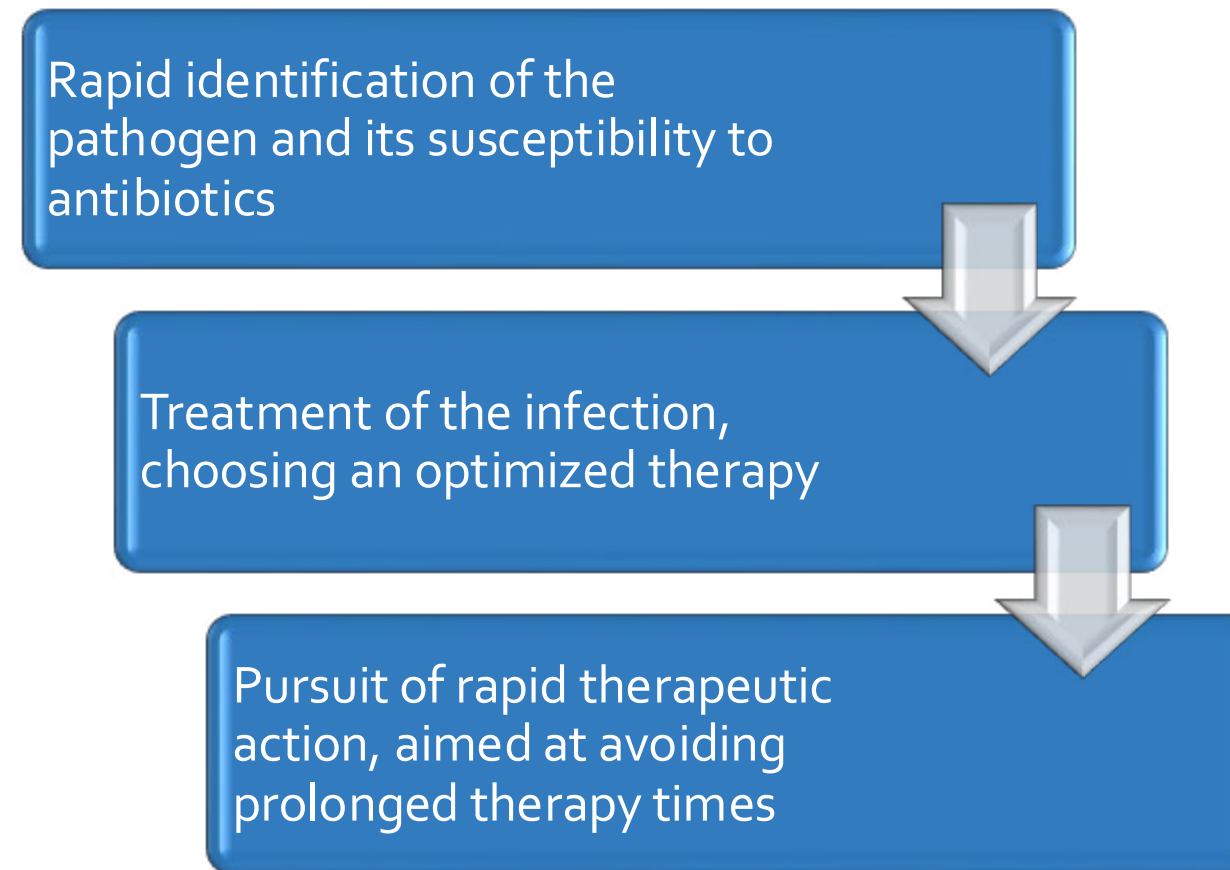
Cheong *et al. Infect Chemother* 2022

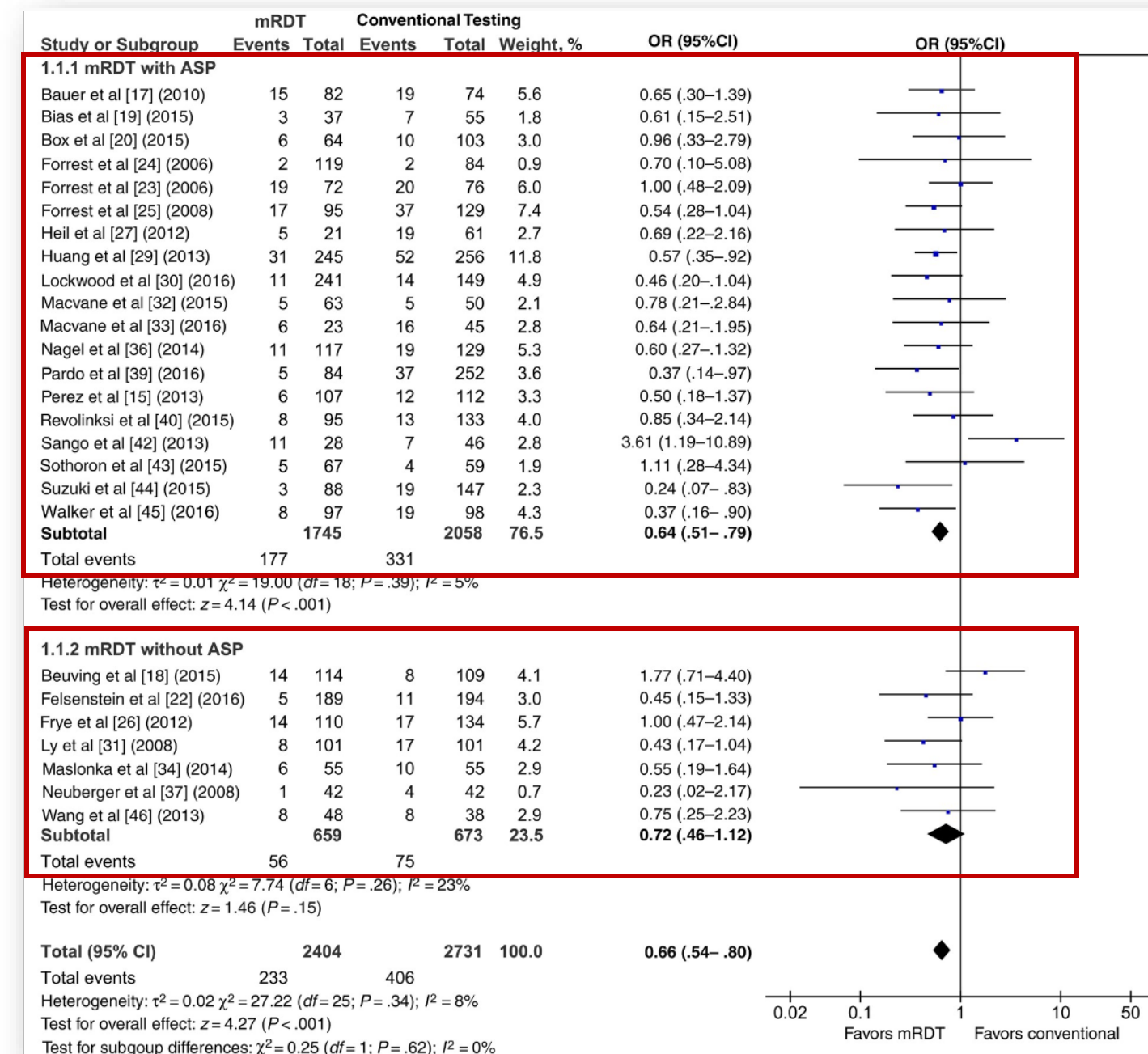
Cortegiani *et al. J Anesth Analg Crit Care* 2023

Multidisciplinary team responsible for implementing antimicrobial stewardship



Appropriate antimicrobial management includes:

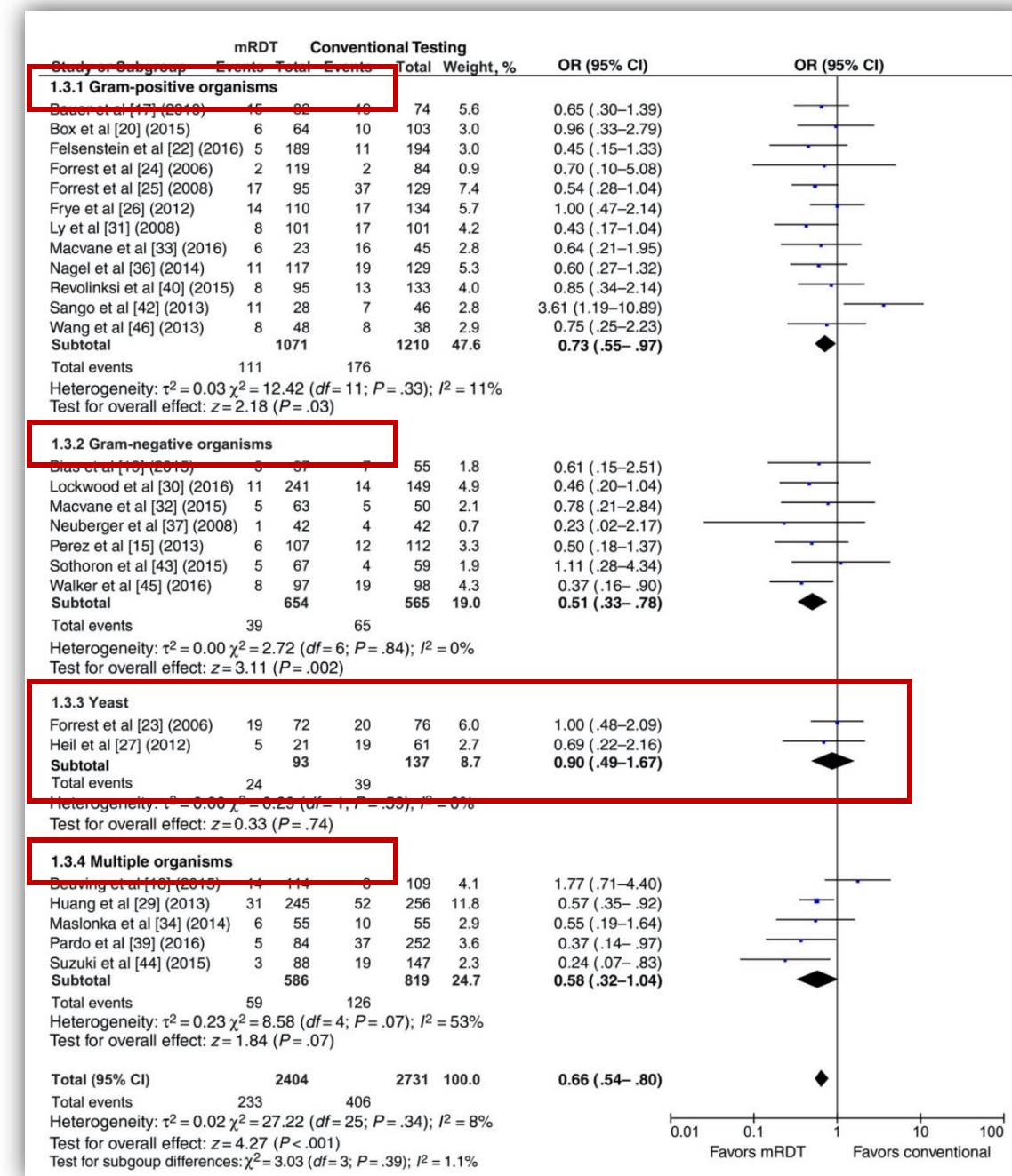




The Effect of Molecular Rapid Diagnostic Testing on Clinical Outcomes in Bloodstream Infections

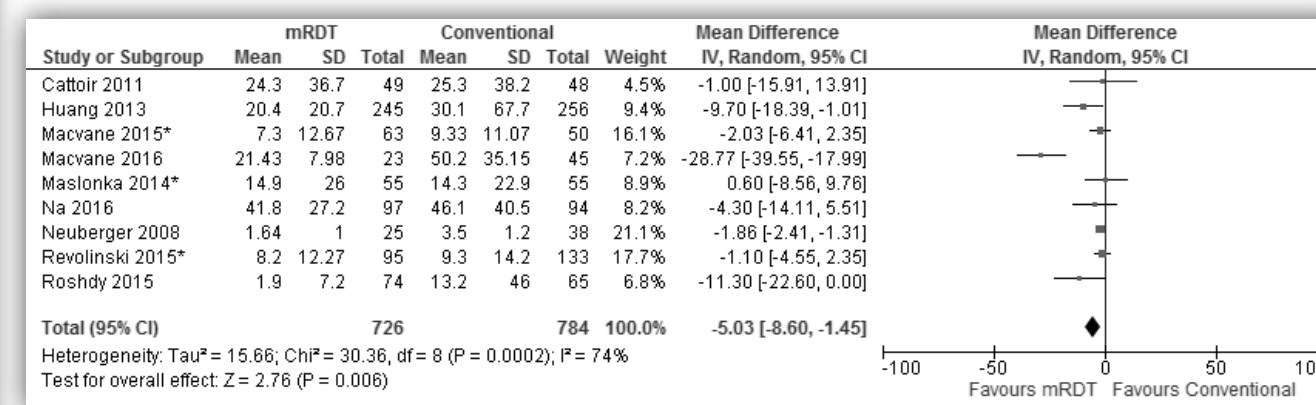
Mortality outcomes with molecular rapid diagnostic testing versus conventional testing in bloodstream infection

Tristan T. Timbrook et al. Clinical Infectious Diseases 2017

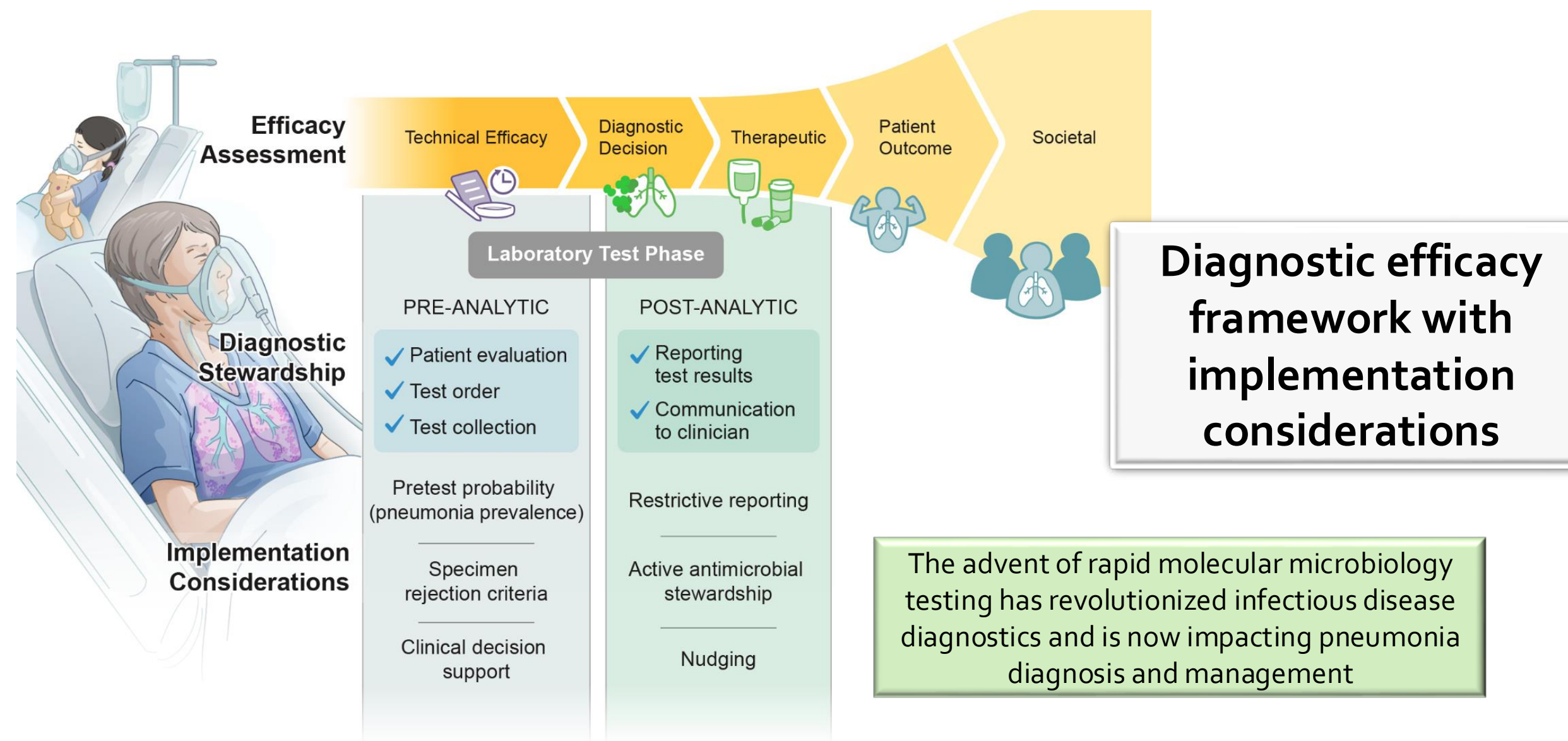


Mortality with molecular rapid diagnostic testing (mRDT) versus conventional testing by organism type in bloodstream infection

Length of stay with mRDT vs conventional testing in BSI



Tristan T. Timbrook et al. Clinical Infectious Diseases 2017



The evolving landscape of microbiology testing yields substantial opportunities for clinical microbiologists and bedside providers to collaborate in developing implementation strategies to maximize the clinical utility of these advanced diagnostics in driving outcomes

Walker AM. et al., Diagnostics (Basel). 2024

- Right test
- Right patient
- Right time

Diagnostic stewardship

Key diagnostic stewardship considerations for implementation of rapid infectious disease diagnostics

1. Is the test appropriate for the clinical setting?

Evaluation of test performance, laboratory feasibility, and cost versus value

2. Will the clinical care of the patient be affected by the test result?

Directing testing toward the right patients



2015 Journal of Clinical Microbiology

Criteria for Reducing Unnecessary Testing for Herpes Simplex Virus, Varicella-Zoster Virus, Cytomegalovirus, and Enterovirus in Cerebrospinal Fluid Samples from Adults

Craig B. Wilen,^a Cynthia L. Monaco,^b Joan Hoppe-Bauer,^c Ronald Jackups, Jr.,^d Robert C. Bucelli,^e Carey-Ann D. Burnham^{f,g}

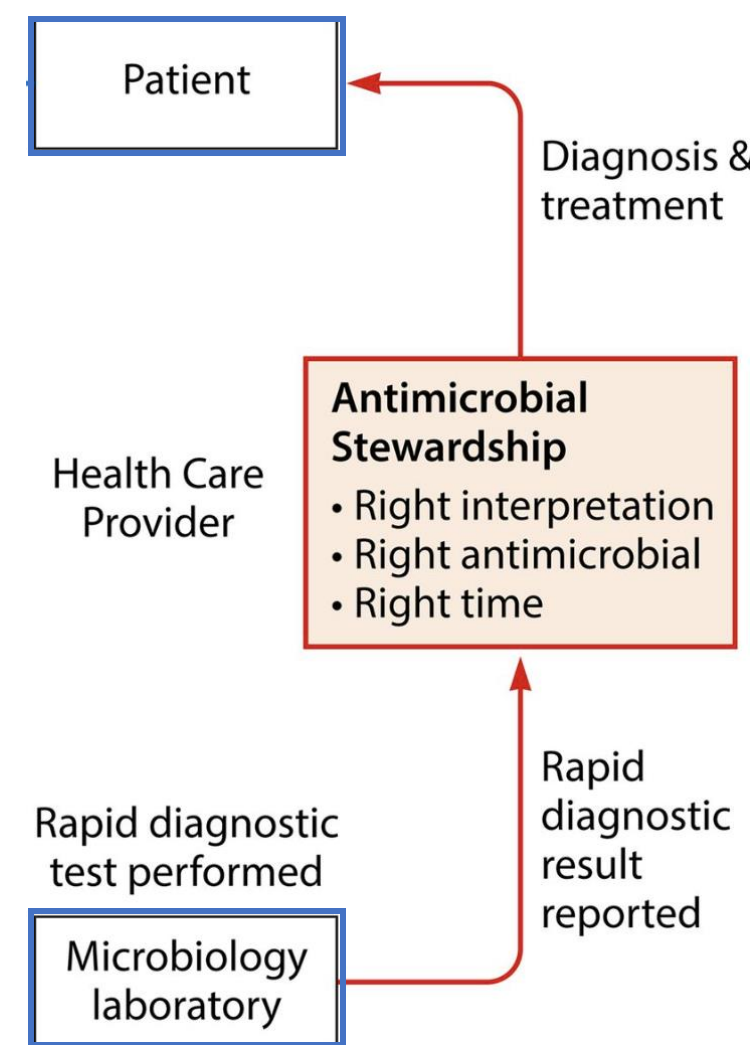
3. Will the result be available in time to optimally affect care?

The time to receive the sample in the clinical microbiology lab and the time to report the results

Messacar K. et al. J Clin Microbiol 2017

Antimicrobial stewardship

Key antimicrobial stewardship considerations for implementation of rapid infectious disease diagnostics



1. Will the clinician understand the test result?

Result report language, selective reporting of relevant results

2. Will the clinician appropriately modify antimicrobials based on test result?

Clinical practice guidelines, EMR-based decision support with result reporting

3. Will the clinician act upon the test result promptly?

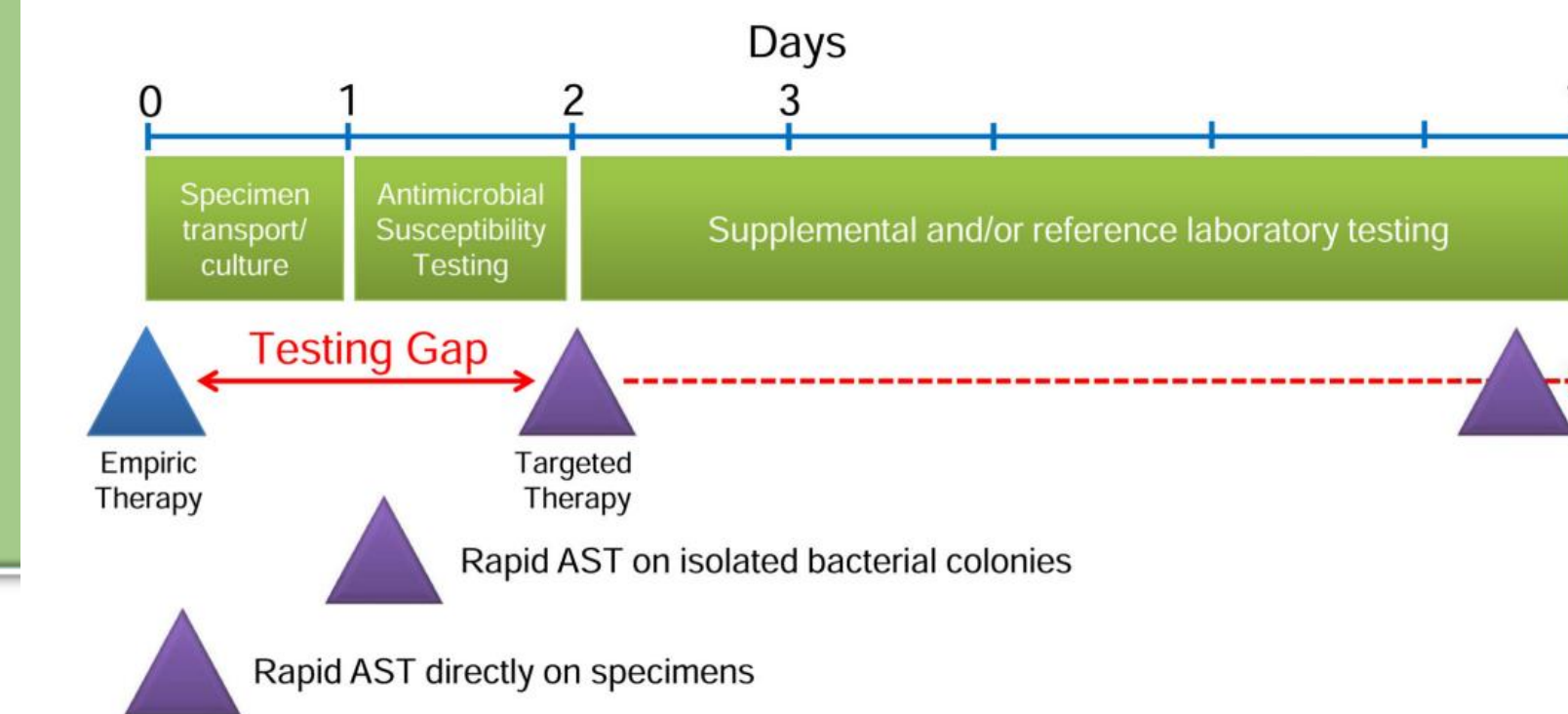
EMR reporting, AS real-time decision support

EMR: electronic medical record
 AS: antimicrobial stewardship

Messacar K. et al. J Clin Microbiol 2017

➤ Time (together with reliability) is one of the most important factors in defining the clinical relevance of the diagnostic tests in microbiology

- Respiratory infections
- Sepsis
- AMR
- Meningitis



If not timely provided to the clinician, in most cases the result is useless

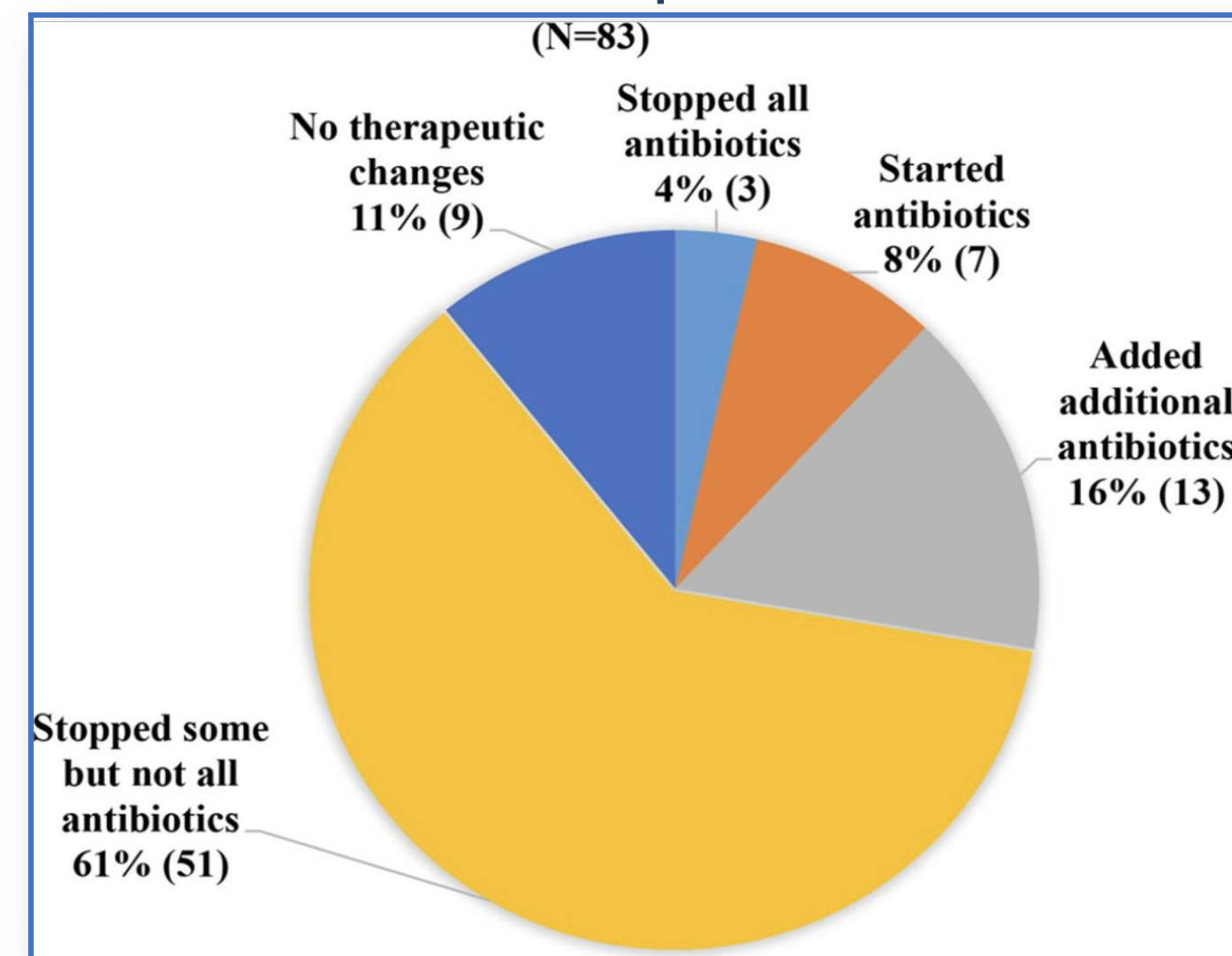
Rapid Susceptibility Testing Methods Kenneth P. Smith, PhD, James E. Kirby, MD
 Clin Lab Med 2019

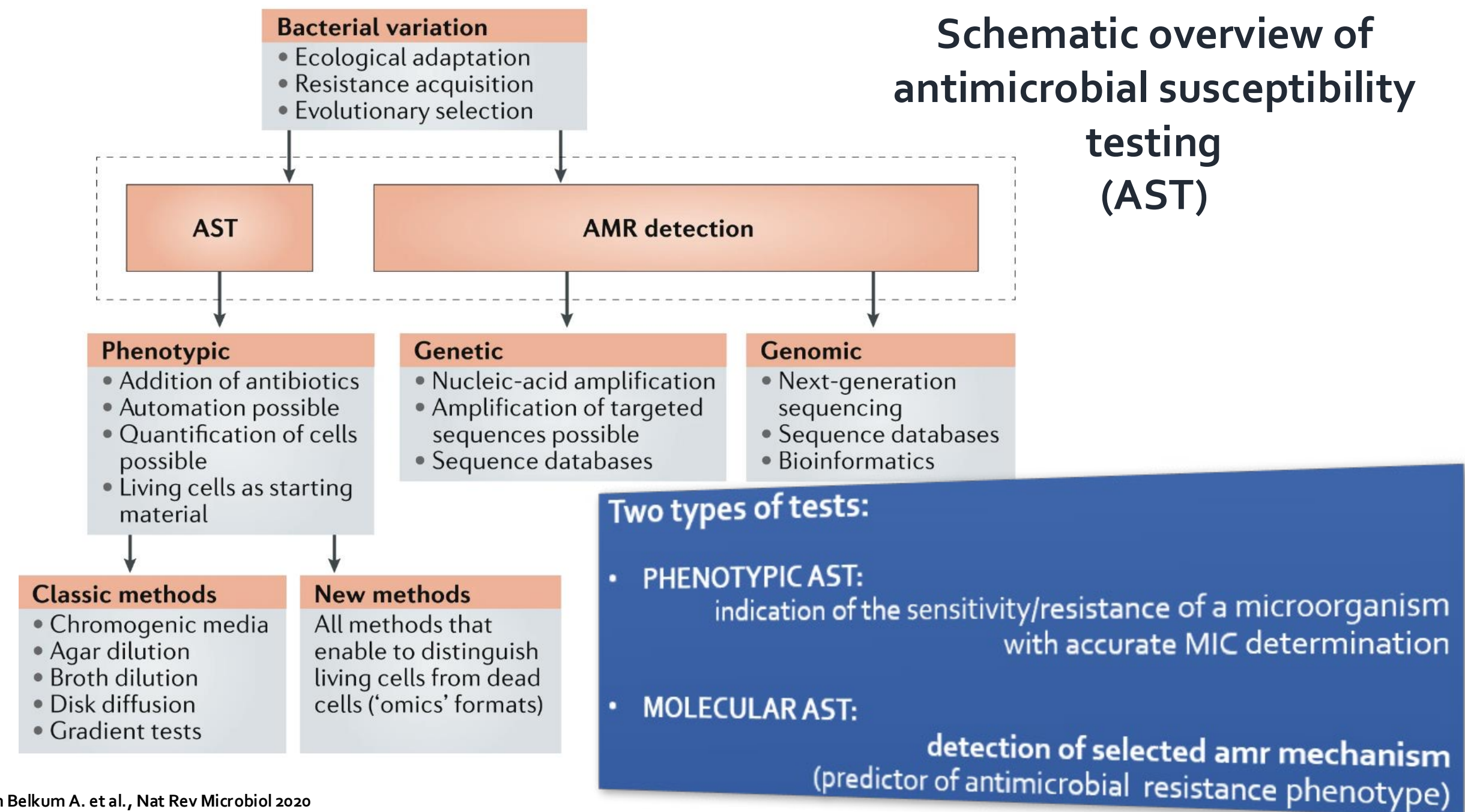
Assessment of Rapid-Blood-Culture-Identification Result Interpretation and Antibiotic Prescribing Practices

Self-reported changes in antibiotic therapy frequently made based on BCID results

BCID results

Blood culture result interpretation





van Belkum A. et al., Nat Rev Microbiol 2020

Ritu Banerjee et al., Frontiers in Medicine 2021

Rapid phenotypic AST methods for testing positive blood cultures

It is mandatory to promptly report accurate MIC values

Test	AST technology	TTR
PhenoTest BC (Accelerate Diagnostics)	Time-lapse imaging of bacterial cells under dark-field microscopy. Morphological and kinetic changes analyzed.	7 h
Alfred (AliFAX)	Light scattering to detect bacterial growth in liquid culture broth.	3–5 h
dRAST (QuantaMatrix)	Time-lapse imaging of bacterial cells on micropatterned plastic microchips.	6 h
Reveal AST (Specific Diagnostics)	Sensor array for volatile organic compounds emitted during microorganism growth.	4.5 h
ASTar (Q-linea)	Time-lapse imaging of bacterial growth in broth.	3–6 h
Fastinov	Flow cytometry applying fluorescent dyes that reveal cell damage during treatment.	80 min
LifeScale (Affinity Biosensors)	Mass measurement using a microcantilever.	4 h

Method	Resistance genes
Alifax Molecular Mouse System GRAM NEG RESIST	bla _{CTX-M} , bla _{SHV} , bla _{CMY-1/MOX} , bla _{CMY-2} , bla _{IIMP} , bla _{NDM} , bla _{KPC} , bla _{VIM} , bla _{OXA-23} , bla _{OXA-48} , mcr-1, mcr-2
BioFire FilmArray Blood Culture Identification 2 (BCID2), Pneumonia plus (PNplus) panels and Joint Infection (JI) Panel	bla _{CTX-M} , bla _{NDM} , bla _{KPC} , bla _{VIM} , bla _{IIMP} , bla _{OXA-48} , mcr-1
Eplex BCID-GN Panel	bla _{CTX-M} , bla _{NDM} , bla _{KPC} , bla _{VIM} , bla _{IIMP} , bla _{OXA-23} , bla _{OXA-48}
Eurospital Sepsis Flow Chip Kit Eurospital MDR Flow Chip Kit	bla _{CTX-M} , bla _{CMY-1/MOX} , bla _{SHV} , bla _{CMY} , bla _{DHA} , bla _{IIMP} , bla _{NDM} , bla _{KPC} , bla _{VIM} , bla _{OXA-23} , bla _{OXA-48} , bla _{NCM/IMI} , bla _{GIM} , bla _{OXA-23} , bla _{OXA-24} , bla _{OXA-51} , bla _{OXA-58} , mcr-1, mcr-2
Unyvero Cartridge (Blood Culture, Hospitalized Pneumonia, Implant & Tissue Infection, Intra-Abdominal Infection, Urinary Tract Infection)	bla _{CTX-M} , bla _{SHV} , bla _{TEM} , bla _{NDM} , bla _{KPC} , bla _{VIM} , bla _{IIMP} , bla _{OXA-23} , bla _{OXA-24/40} , bla _{OXA-48} , bla _{OXA-58}
Verigene Gram-Negative Blood Culture test	bla _{CTX-M} , bla _{NDM} , bla _{KPC} , bla _{VIM} , bla _{IIMP} , bla _{OXA-48}
Cefide Xpert Carba-R	bla _{NDM} , bla _{KPC} , bla _{VIM} , bla _{IIMP} , bla _{OXA-48}
Eazyplex® SuperBug CRE	bla _{NDM} , bla _{KPC} , bla _{VIM} , bla _{OXA-48} , bla _{OXA181} , bla _{CTX-M-1} , bla _{CTX-M-9}
Eazyplex® SuperBug complete C	bla _{CTX-M} , bla _{NDM} , bla _{KPC} , bla _{VIM} , bla _{IIMP} , bla _{OXA-48} , bla _{OXA181}
Eazyplex® SuperBug AmpC	bla _{ACC} , bla _{DHA} , bla _{CMYII} , bla _{MOX}
Hologic Novodiag® CarbaR+ Assay	bla _{NDM} , bla _{KPC} , bla _{VIM} , bla _{IIMP} , bla _{OXA-48/181} , bla _{OXA23} , bla _{OXA24} , bla _{OXA51} , bla _{OXA58} , mcr-1

Molecular-genetic techniques for detection of gram negative resistance genes

Genetic and genomic detection of MRSA

Molecular Method Used	Principle of the Method	On Culture/on Clinical Sample	TAT ¹	Brief Advantages/Disadvantages +/-	No. and Type of <i>S. aureus</i> Analyzed ²	Major Diagnostic Performance ³
Xpert® SA Nasal Complete (Cepheid)	Real-time PCR for <i>mecA/C</i> , <i>spa</i> and <i>SCCmec-orfX</i>	Nasal samples	3 h	+ Clinical outcomes analyzed - 56 invalid results not further analyzed	10 MRSA in 605 nasal samples	Sensitivity 100% Specificity 98.8% PPV 58.8% NPV 100% TAT 41 h shorter
Xpert® MRSA/SA BC Assay (Cepheid)	Real-time PCR for <i>mecA/C</i> , <i>spa</i> and <i>SCCmec-orfX</i>	Positive blood cultures	85 m	- Unusual reference method	27 MRSA in 500 nasal samples	Sensitivity 51.8% Specificity 100%
			1.7 h	+ Clinical outcomes analyzed - More resistant isolates needed	1 MRSA 38 MSSA in 264 blood cultures	Sensitivity 100% specificity 100% TAT 24 h shorter, earlier changes in patient management
Xpert® MRSA/SA SSTI (Cepheid)	Real-time PCR for <i>mecA/C</i> , <i>spa</i> and <i>SCCmec-orfX</i>	BAL samples	n.a.	+ Clinical outcomes analyzed - Sensitivity/specificity not calculated	37 MRSA 64 MSSA	Time to optimal therapy 20 h shorter, duration of vancomycin therapy 18 h shorter
			68 m	+ Clinical outcomes analyzed - Method not validated in BAL samples	23 MRSA 25 MSSA in 247 BAL samples	Sensitivity 95.7% specificity 98.2% Time of linezolid/vancomycin treatment 40 h shorter
Hologic Panther Fusion® MRSA	PCR and Invader chemistries for <i>mecA/C</i> , <i>gap</i> and <i>SCCmec-orfX</i>	Nasal samples	<3 h	+ Can analyze 350 samples in 8 h - Need comparison with a similar method	30 MRSA 112 MSSA in 434 nasal swabs	Sensitivity 86.7%, specificity 98.8%, CA 97.9%
MRSA/SA ELITE MGB assay (ELITechGroup)	Real time PCR for <i>mecA/C</i> and a <i>S. aureus</i> specific sequence	Sputum, tracheal aspirate, BAL	<3 h	+ Accurate - Do not target <i>SCCmec-orfX</i> junction	23 MRSA 60 MSSA in 113 respiratory samples	Sensitivity 95.7% specificity 96.7% PPV 91.7% NPV 98.3%
Unyvero HPN Application	Multiple PCRs	BAL fluids	5 h	+ Detect 21 species and 19 resistance genes; mixed cultures detection - More resistant isolates needed	2 MRSA 1 MSSA in 84 BAL fluids	Sensitivity 100% specificity 98.7%

Molecular assays for detection of VRE

Method	Resistance genes
Alifax Molecular Mouse System GRAM POS RESIST	vanA, vanB
BioFire FilmArray Blood Culture Identification 2 (BCID2), Pneumonia plus (PNplus) panels and Joint Infection (JI) Panel	vanA/B
Eplex BCID-GN Panel	vanA, vanB
Eurospital Sepsis Flow Chip Kit Eurospital MDR Flow Chip Kit	vanA, vanB
Unyvero Cartridge (Blood Culture, Hospitalized Pneumonia, Implant & Tissue Infection, Intra-Abdominal Infection, Urinary Tract Infection)	vanA, vanB
Verigene Gram-Positive Blood Culture test	vanA, vanB
Eazyplex VRE	vanA, vanB

Antibiogramma cumulativo – materiali non invasivi anno 2023



3	GRAM - (Materiali non Invasivi)	Total	Ampicillina	Amoxicillina/ clavulanato	Piperacillina/ tazobactam	Cefotaxime	Ceftazidime	Cefepime	Meropenem	Imipenem	Amikacina	Gentamicina	Ciprofloxacina	Trimetoprim/ sulfametossazolo	Colistina	Ceftazidime/ avibactam	Ceftolozane/ tazobactam
	<i>Acinetobacter baumannii</i>	37	R	R		R			88.6%	83.3%			80.6%	97.0%			
	<i>Enterobacter cloacae</i>	163	R	R	74.6%	69.67%	71.53%	81.64%	93.78	92.80%	96.9%(1)	93.9%(1)	90.7%	81.3%		92.8%	81.7%
	<i>Escherichia coli</i>	445			86.9%	79.9%	79.7%	84.3%	99.3%	98.3%	97.3%(1)	84.0%(1)	72.7%	59.0%		100.0%	93.1%
	<i>Klebsiella pneumoniae</i>	278	R	67.9%	62.2%	50.0%	50.0%	50.7%	92.4%	91.1%	99.1%(1)	71.9%(1)	52.2%	50.0%		98.7%	78.1%
	<i>Proteus mirabilis</i>	79		77.8%	100.0%	93.0%	69.0%	96.1%	97.4%	R	100.0%(1)	96.0%(1)	75.3%	56.9%	R	100.0%	74.1%
	<i>Pseudomonas aeruginosa</i>	482	R	R	70.3%	R	76.5%	76.6%	79.3%	77.4%	98.1%(1)		81.0%	R		84.0%	81.7%
	<i>Serratia marcescens</i>	90	R	R		84.5%	87.5%	96.6%	93.3%		100.0%(1)	100.0%(1)	98.8%	98.8%	R	97.7%	90.7%
	<i>Stenotrophomonas maltophilia</i>	149	R	R	R	R			R	R	R	R		99.1%			

4	GRAM + (Materiali non Invasivi)	Total	Ampicillina	Penicillina	Oxacillina	linezolid	Daptomicina	Imipenem	Eritromicina	Levofloxacina	Teicoplanina	Vancomicina	Trimetoprim/ sulfametossazolo	Gentamicina	Tetraciclina	Quinupristin/ dalopristin
	<i>Enterococcus faecalis</i>	259	100.0%			100.0%		100.0%			100.0%	100.0%		R		R
	<i>Enterococcus faecium</i>	131	5.3%			100.0%		5.3%			73.8%	73.8%		R		
	<i>Staphylococcus aureus</i>	784		23.2%	79.9%	100.0%	100.0%		55.4%	80.5%	100.0%	100.0%	97.3%	()	89.1%	100.0%
	<i>Staphylococcus epidermidis</i>	85			12.9%	100.0%	100.0%		20.2%	32.1%	98.7%	100.0%	51.9%	()	34.1%	100.0%
	<i>Staphylococcus haemolyticus</i>	44			7.0%	100.0%	100.0%		4.5%	7.0%	67.5%	100.0%	23.3%	()	22.7%	100.0%
	<i>Staphylococcus hominis</i>	12			33.3%	100.0%	100.0%		33.3%	66.7%	100.0%	100.0%	50.0%	()	75.0%	100.0%

Antibiogramma cumulativo - Percentuale di microrganismi sensibili isolati da materiali non invasivi

NOTE: considerati solo campioni da tratto urinario

Legenda:

	>= 90% S
	70% - 89% S
	< 70% S
	Intrinsecamente Resistente
	Breakpoints in brackets

Antibiogramma cumulativo – materiali invasivi anno 2023



1		Total	Ampicillina	Amoxicillina/ clavulanato	Piperacillina/ tazobactam	Cefotaxime	Ceftazidime	Cefepime	Meropenem	Imipenem	Amikacina	Gentamicina	Ciprofloxacina	Trimetoprim/ sulfametossazolo	Colistina	Ceftazidime/ avibactam	Ceftolozane/ tazobactam
GRAM - (Materiali Invasivi)																	
	<i>Acinetobacter baumannii</i>	6	R	R		R			33.3%	20.0%	()	()	20.0%	66.0%			
	<i>Enterobacter cloacae</i>	45	R	R	80.9%	66.7%	60.0%	79.1%	93.3%	90.9%	()	()	77.8%	82.0%		91.1%	82.2%
	<i>Escherichia coli</i>	37			83.8%	75.7%	73.0%	78.4%	100.0%	100.0%	()	()	66.7%	58.3%		100.0%	94.6%
	<i>Klebsiella pneumoniae</i>	51	R	53.6%	46.9%	40.8%	41.6%	46.5%	80.4%	84.3%	()	()	41.2%	38.0%		100.0%	70.0%
	<i>Pseudomonas aeruginosa</i>	37	R	R	59.5%	R	67.6%	67.7%	59.5%	63.9%	()	()	85.7%	R		86.1%	88.9%
	<i>Proteus mirabilis</i>	4		66.7%	100.0%	100.0%	100.0%	100.0%	100.0%	R	()	()	75.0%	50.0%	R	100.0%	75.0%
	<i>Serratia marcescens</i>	12	R	R		81.8%	100.0%	100.0%	100.0%		()	()	83.3%	100.0%	R	100.0%	100.0%
	<i>Stenotrophomonas maltophilia</i>	9	R	R	R	R			R	R	R	R	100.0%				
GRAM + (Materiali Invasivi)																	
2		Total	Ampicillina	Penicillina	Oxacillina	linezolid	Daptomicina	Imipenem	Eritromicina	Levofloxacina	Teicoplanina	Vancomicina	Trimetoprim/ sulfametossazolo	Gentamicina	Tetraciclina	Quinupristin/ dalpristin	
	<i>Enterococcus faecalis</i>	35	100.0%			100.0%		100.0%			100.0%	100.0%		R		R	
	<i>Enterococcus faecium</i>	23	8.7%			100.0%		8.7%			73.9%	73.9%		R			
	<i>Staphylococcus aureus</i>	80		16.3%	75.0%	100.0%	100.0%		56.3%	78.8%	100.0%	100.0%	100.0%	()	87.5%	100.0%	
	<i>Staphylococcus epidermidis</i>	298			14.8%	100.0%	100.0%		28.5%	41.4%	95.1%	100.0%	63.3%	()	34.2%	100.0%	
	<i>Staphylococcus haemolyticus</i>	73			17.8%	100.0%	100.0%		8.2%	23.3%	97.1%	100.0%	39.7%	()	26.1%	100.0%	
	<i>Staphylococcus hominis</i>	142			34.8%	100.0%	100.0%		21.8%	72.3%	100.0%	100.0%	63.8%	()	54.2%	100.0%	

Antibiogramma cumulativo - Percentuale di microrganismi sensibili isolati da materiali invasivi

Legenda:

	>= 90% S
	70% - 89% S
	< 70% S
	Intrinsecamente Resistente
	Breakpoints in brackets

The 24-h clinical microbiology service is essential for patient management

Joseph M Blondeau^{*1,2} & Evgeny A Idelevich³



“In summary, optimal patient care requires access to necessary laboratory testing including clinical microbiology. A rethinking of hours of operation is required to shorten time to accurate result reporting.”

Blondeau JM, Idelevich EA. Future Microbiol. 2018


Concluding remarks

Emerging antimicrobial resistance **makes empiric therapy unreliable**. Therefore, rapid antimicrobial susceptibility testing will provide early, definitive therapeutic guidance to optimize patient outcome


Genotypic rapid AST methods are fast but can only identify what we know about

Phenotypic rapid AST methods provide a nuanced integrated assessment of resistance that can be used to pick the most active therapy

The combined use of new technologies allows a fast diagnostic approach which shows a positive impact on early targeted therapy



The key elements to be acquired to protect frail patients, are: speed, accuracy of diagnosis, therapeutic appropriateness of new antibiotics, and the need for multidisciplinary task forces



Agree with the microbiologist on the most appropriate fast strategy to adopt...

*Thank you
 for your attention*

PAOLA BERNASCHI

UOC Microbiologia e Diagnostica di Immunologia
 Ospedale Pediatrico Bambino Gesù (IRCCS)
 Roma

Microbiology Staff OPBG

Prof. Carlo Federico Perno

- Agosta Marilena
- Argentieri Marta
- Elena Chaiter
- Cortazzo Venere
- Fox Valeria
- Lucignano Barbara
- Mancinelli Livia
- Onori Manuela
- Pansani Laura
- Rossitto Martina
- Sisto Annamaria
- Tuccio Guarna Assanti Vanessa
- Vrenna Gianluca

- Angelaccio Anna
- Cetra Vittoria
- De Santis Maria Luisa
- Di Leva Francesca
- D'Urbano Teresa
- Ferri Giulia
- Foglietta Gianluca
- Parlavecchio Carmen
- Tredici Silvia
- Zullino Ilaria

"good microbiology is clinically relevant microbiology"

